Survival, growth, and maturation of secondary follicles from prepubertal, young, and older adult rhesus monkeys during encapsulated three-dimensional culture: effects of gonadotropins and insulin

Jing Xu1, Marcelo P Bernuci1, Maralee S Lawson1, Richard R Yeoman1, Thomas E Fisher1,2, Mary B Zelinski1,2 and Richard L Stouffer1,2

1Division of Reproductive Sciences, Oregon National Primate Research Center, Oregon Health & Science University, 505 Northwest 185th Avenue, Beaverton, Oregon 97006, USA and 2Department of Obstetrics and Gynecology, Oregon Health & Science University, 3181 SW Sam Jackson Park Road, Portland, Oregon 97239, USA

Correspondence should be addressed to M B Zelinski at Division of Reproductive Sciences, Department of Obstetrics and Gynecology, Oregon National Primate Research Center, Oregon Health & Science University; Email: zelinski@ohsu.edu

Abstract

A three-dimensional culture system supports the development of primate preantral follicles to the antral stage with appreciable steroid production. This study assessed i) whether in vitro developmental competence of follicles is age dependent, ii) the role of gonadotropins and insulin in supporting folliculogenesis, and iii) anti-Müllerian hormone (AMH) and vascular endothelial growth factor (VEGF) production by growing follicles. Ovaries were obtained from prepubertal, young, and older adult rhesus macaques. Secondary follicles were encapsulated into alginate beads and cultured individually for 40 days in media containing 0.05 or 5 μg/ml insulin, with or without recombinant human (rh) FSH (500 mIU/ml). No follicles survived in the culture without rhFSH. In the presence of rhFSH, survival was lower for follicles from older animals, whereas growth, i.e. follicle diameter, was less by day 40 for follicles from prepubertal animals. The surviving follicles were categorized as no-grow (NG; ≤250 μm), slow-grow (SG; 250–500 μm), and fast-grow (FG; ≥500 μm) according to their diameters. SG follicles cultured with 5 μg/ml insulin produced more ovarian steroids than those cultured with 0.05 μg/ml insulin by week 5. SG and FG follicles produced more AMH and VEGF than the NG, and levels peaked at weeks 2 and 5 respectively. After 100 ng/ml rh chorionic gonadotropin treatment for 34 h, more healthy oocytes were retrieved from young adults whose follicles were cultured with 5 μg/ml insulin. This culture system offers an opportunity to characterize the endocrine and paracrine function of primate follicles that influence follicle growth and oocyte maturation.

Reproduction (2010) 140 685–697

Introduction

Early follicular development is a dynamic process that is regulated by complex interactions between gonadotropins and local factors (Gougeon 1996). Although progress in understanding early folliculogenesis has been made, particularly in mice through gene manipulation (Matzuk 2000, Drummond 2006), the regulation and dynamics of primate folliculogenesis, aside from phenotypic analysis in human (Chand et al. 2010, Ewens et al. 2010), remain poorly understood. Recently, feasibility of using tissue engineering principles to develop a three-dimensional (3D) matrix that supports follicle development was successfully demonstrated in mice (Xu et al. 2006, West et al. 2007). This 3D approach yielded antral follicles with mature oocytes that, when inseminated in vitro, fertilized and, following embryo transfer, produced live offspring (Xu et al. 2006).

Applying encapsulated 3D follicle culture to the primate, it was possible to grow rhesus monkey preantral follicles through the small antral stage with appreciable levels of ovarian steroid production (Xu et al. 2009a). This technique provides a valuable in vitro model to study the process and regulation of folliculogenesis in intact individual follicles in primates. However, further advances are needed to develop an adequate milieu for macaque follicular growth and differentiation that provides oocytes competent for fertilization and development. The major challenges include i) the length of time for a follicle to grow from preantral to the small antral stage at which it can be selected for final maturation, which is estimated to be 65 days in vivo (Gougeon 1998), and ii) the large size of a mature preovulatory follicle with a fertilizable oocyte, which is 3–6 mm in diameter in macaques.
Although full knowledge of the milieu for follicle growth in vivo is still lacking, there is evidence that the adequate levels of certain hormones are essential for the growth of healthy follicles in vitro (Picton et al. 2008). The pituitary gonadotropin, FSH, may promote, if not be essential for, follicle growth in culture systems. For example, FSH administration not only maintained the morphological integrity of caprine preantral follicles, but also stimulated the activation of primordial follicles and the growth of activated follicles in the culture (Matos et al. 2007). In 3D matrix systems, FSH increased the diameter of mouse (Xu et al. 2006), monkey (Xu et al. 2009a), and human (Xu et al. 2009b) follicles in vitro. LH may also enable primary and secondary follicles to respond to later LH-dependent growth, by inducing early differentiation of the theca cells, during follicle culture in mice (Cortrvindt et al. 1998, Wu et al. 2000). At later stages of culture, monkey follicles in the media supplemented with LH tended to produce more estradiol (E2), which may be attributed to increased presence of theca cell (Xu et al. 2009a). There is also an increasing body of evidence that insulin may stimulate follicular growth in vitro (Wright et al. 1999). Insulin, as a survival factor, reduces the proportion of atretic follicles in early stages of human follicle culture, hence improving the overall viability of the follicles (Louhio et al. 2000).

It is also likely that paracrine factors modulate preantral and antral follicle development in primates (Albertini et al. 2001, Eppig 2001). Growing evidence suggests that anti-Müllerian hormone (AMH) plays a key role in the early stages of folliculogenesis (Durlinger et al. 2002, Carlsson et al. 2006). AMH could affect early stages of human follicular development by enhancing recruitment, survival, and/or growth during ovarian tissue culture (Schmidt et al. 2005). Also, the angiogenesis process is a fundamental activity in the ovary, specifically during follicular growth, atresia, ovulation, and luteinization. Angiogenic factors produced by the follicle, such as vascular endothelial growth factor (VEGF), may promote vascular changes during follicle growth (Hazzard & Stouffer 2000). However, they may also have other roles, as suggested by evidence that VEGF maintains follicular ultrastructural integrity and promotes the growth of caprine preantral follicles during ovarian tissue culture (Bruno et al. 2009).

Thus, AMH and VEGF may be valuable markers of follicle maturation and function. However, knowledge surrounding the dynamics of AMH and VEGF production and regulation in the growing follicles is unknown, especially in primates.

Using encapsulated 3D culture of nonhuman primate preantral follicles, this study assessed whether the developmental competence of follicles was dependent upon maternal age, and evaluated the role of gonadotropins, FSH alone or with sequential addition of LH, and insulin to support folliculogenesis. Additionally, studies were conducted to investigate the ovarian steroids (androstenedione, A4; E2; and progesterone, P4) and paracrine factors (AMH and VEGF) produced by individual cultured follicles that may influence follicle growth and health.

**Results**

**Follicle survival and growth**

Total and average number of secondary follicles isolated from different age groups of animal and the follicle distribution in different culture conditions are summarized in Table 1. Follicle survival could be identified clearly with viable follicles exhibiting an intact basement membrane versus atretic follicles having dark granulosa cells and denuded oocytes (Xu et al. 2009a). By culture week 2, all follicles cultured in the absence of FSH underwent atresia regardless of animal age and culture condition, whereas more than 50% of the follicles cultured with FSH survived.

For follicles cultured in the presence of FSH and high-dose insulin, there was no difference in survival rate between prepubertal (59±8%) and young adult (55±11%) animals by culture week 5. However, the survival rate was less (P<0.05) for follicles from older adult animals (15±5%) by culture week 5 than those from prepubertals and young adults. The lower insulin dose and LH administration had no effect on follicle survival for animals in all the three age groups (data not shown).

At the beginning of the culture, follicle diameters did not differ among the age groups and culture conditions (data not shown). However, during culture, three distinct cohorts of follicles were observed based on their growth rate. For example, for follicles cultured with FSH and high-dose insulin (Fig. 1A), a cohort remained similar in size to the initial secondary follicles without significant change in diameters through 5 weeks of culture, ranging from 148±3 to 195±40 µm in young adult animals.

<table>
<thead>
<tr>
<th>Age group</th>
<th>Total yield (mean±S.E.M.)</th>
<th>Without FSH</th>
<th>With FSH</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>5 µg/ml insulin</td>
<td>0.05 µg/ml insulin</td>
</tr>
<tr>
<td>Prepubertal</td>
<td>204 (68±4)</td>
<td>24</td>
<td>12</td>
</tr>
<tr>
<td>Young adult</td>
<td>324 (100±0)</td>
<td>24</td>
<td>12</td>
</tr>
<tr>
<td>Older adult</td>
<td>360 (120±7)</td>
<td>24</td>
<td>12</td>
</tr>
</tbody>
</table>

Table 1 Total and average yield of follicles from animals of different age groups and their assignment to experimental culture conditions.

---

(Fig. 1B), and were termed ‘no-grow (NG)’ follicles. Another cohort doubled \((P<0.05)\) their diameters, from 158±8 to 376±25 \(\mu\)m in young adult animals (Fig. 1C), and were termed ‘slow-grow (SG)’ follicles. Finally, another group of follicles increased \((P<0.05)\) their diameters by a minimum of threefold, from 165±8 to 667±58 \(\mu\)m in young adult animals (Fig. 1D), and were termed ‘fast-grow (FG)’ follicles. The NG follicles from prepubertal and older, as well as from young adults were evident in all ovarian samples (Fig. 1B). The SG pattern also appeared in follicles from both young and older adult animals (Fig. 1C), but the SG follicles from young adults reached larger \((P<0.05)\) diameters by culture week 5 than those from the prepubertal group (Fig. 1C). The FG follicles were only collected from young adult animals and represented 43% of the total cohort; whereas 9% were NG and 48% were SG follicles. The percentage of NG and SG follicles were 14 and 86% respectively for the prepubertal group. In contrast, there was a greater proportion of NG (69%) and a smaller proportion of SG follicles (31%) collected from the older adult group (Fig. 1A). An antral cavity was evident within 3 weeks of culture for all the SG and FG follicles.

The lower dose of insulin did not alter the growth rate of the NG follicles (Fig. 2A) and SG follicles (Fig. 2B) from young adult animals, as well as from the prepubertal and older adult monkeys (data not shown).

### Figure 1
Macaque follicle survival and growth in media containing FSH and high-dose insulin, without LH supplementation at day 30. Number of three cohorts of surviving follicles from different age groups at week 5 of culture (panel A) and the growth patterns for the no-grow (panel B), slow-grow (panel C), and fast-grow (panel D) follicles from prepubertal \((n=3)\), young \((n=3)\), and older \((n=3)\) adult animals during culture. Significant differences over time (lowercase) or between the age groups (uppercase) are indicated by different letters \((P<0.05)\). Data are presented as the mean ± S.E.M.

### Figure 2
Insulin effect on patterns of macaque follicle growth. Growth patterns of no-grow (panel A), slow-grow (panel B), and fast-grow (panel C) follicles from young adult animals \((n=3)\) during culture in media containing FSH and either low- or high-dose insulin, without LH supplementation at day 30. Significant differences over time (lowercase) or between the insulin dosage groups (uppercase) are indicated by different letters \((P<0.05)\). Data are presented as the mean ± S.E.M.

However, even though the FG follicles maintained a similar growth rate between the low- and high-dose insulin-treated groups for the first 3 weeks of culture, the follicle diameters became larger \((P<0.05)\) in the high-dose insulin cultures than in the low-dose cultures on weeks 4 and 5 (Fig. 2C).

LH supplementation at culture day 30 did not promote further growth of the follicles regardless of animal age and culture condition (data not shown).

### Oocyte maturation and diameter
Following exposure of antral follicles (SG and FG) to recombinant human chorionic gonadotropin (rhCG), healthy, as well as degenerate (dark and condensed cytoplasm), oocytes were obtained during retrieval. There were also follicles that did not yield oocytes.
Despite rhCG treatment, all the healthy oocytes remained at the germinal vesicle (GV)-intact stage at retrieval (Table 2), and no oocytes matured to metaphase I (MI) or MII stage during 48 h in vitro after removal from the follicles. Oocytes retrieved from follicles under all the culture conditions were combined for analysis of oocyte maturation among the different age groups. Of the 20 oocytes retrieved from the young adult group, 60% were GV-intact oocytes. Furthermore, from the FG follicles in this group, 86% of the oocytes retrieved were at the GV-intact stage. In contrast, 38% of the oocytes retrieved from the prepubertal animals were at the GV-intact stage, and only two GV-intact oocytes were retrieved from the older adult group.

The oocyte diameters of the FG, but not the SG, follicles from young adults were larger (P<0.05) than those from the prepubertal group (Table 2). Oocyte retrieval from follicles of young adult animals was analyzed among the different culture conditions. The highest percentage of GV-intact oocytes, 70%, were observed from the follicles cultured with FSH, high-dose insulin, and LH. However, there was no significant difference in oocyte diameters among the different culture conditions (Table 3).

**Follicular steroids**

Since media steroid levels from the NG and SG follicles of prepubertal and older adult animals showed similar trends as those from young adults over the 5-week culture period (data not shown), only the data and analyses from the young adult group are presented in graphical form. For the NG follicles cultured within FSH and either low- or high-dose insulin, E2 levels remained at baseline throughout the culture period (Fig. 3A). For the SG (Fig. 3B) and FG (Fig. 3C) follicles cultured with FSH and high-dose insulin, E2 concentrations started rising by weeks 2–3 and were higher (P<0.05) by weeks 3–4 than those observed in the beginning of culture. E2 levels for the SG follicles cultured with FSH and low-dose insulin stayed at baseline and were lower (P<0.05) than those of the high-dose insulin-treated follicles at culture weeks 4 and 5 (Fig. 3B). There were no samples available for the FG follicles cultured with low-dose insulin. Similar patterns were also observed for A4 and P4 levels during the culture interval (e.g. SG follicles, Fig. 4A and E).

In cultures of the SG follicles, levels of A4 (Fig. 4A), E2 (Fig. 4C), and P4 (Fig. 4E) did not increase between weeks 4 and 5 in the presence of FSH and high-dose insulin. However, the addition of LH at culture day 30 increased (P<0.05) A4 (Fig. 4B), E2 (Fig. 4D), and P4 (Fig. 4F) production between pre-LH (week 4) and post-LH (week 5) exposure. In contrast, LH treatment had no effect on media steroid concentrations for the SG follicles cultured with low-dose insulin, which remained at baseline levels throughout the 5 weeks of culture (Fig. 4A–F).

In contrast to the SG follicles, the media levels of A4 (Fig. 5A), E2 (Fig. 5B), and P4 (Fig. 5C) of the FG follicles increased (P<0.05) between 4 and 5 weeks of culture in the presence of FSH and high-dose insulin. Moreover, LH supplementation at culture day 30 had no effect on the pattern or levels of steroids during culture week 5 (Fig. 5A–C).

Figure 6 summarizes steroid levels produced by the SG follicles from monkeys of various ages during culture week 5. Follicles from young adult animals displayed
Anti-Müllerian hormone

When follicles from young adult monkeys were cultured with FSH and high-dose insulin, the media AMH levels produced by the NG follicles did not change throughout the 4 weeks of culture (Fig. 7). In contrast, AMH levels of the SG and FG follicles peaked (P<0.05) at week 2 and then declined (P<0.05) to basal levels by week 3 (Fig. 7). Although diameters of the NG, SG, and FG follicles were not different on week 1 (data not shown), the levels of AMH produced by the SG and FG follicles were higher (P<0.05) than those produced by the NG follicles. Also, AMH levels during week 2 were distinct (P<0.05) among all the three follicle categories (Fig. 7). By week 3, all cultured follicles had basal levels of AMH that remained low even after LH treatment in week 5 (data not shown). Similar patterns were obtained for AMH concentrations throughout the culture period regardless of animal age and culture condition (data not shown).

Vascular endothelial growth factor

When follicles from young adult monkeys were cultured with FSH and high-dose insulin, the media VEGF levels produced by the NG follicles did not change throughout the 5 weeks of culture (Fig. 8). In contrast, VEGF levels of the SG and FG follicles increased markedly (P<0.05)
at weeks 4 and 5 (Fig. 8). Also, VEGF levels during weeks 4 and 5 were distinct \( (P<0.05) \) among all the three follicle categories. By culture week 5, the presence of LH did not alter VEGF concentrations for the SG or FG follicles (data not shown). Similar patterns were obtained for VEGF concentrations throughout 5 weeks of culture regardless of animal age and culture condition (data not shown).

**Discussion**

As described previously \( (Xu \ et \ al. \ 2009a) \), an alginate-based 3D culture system will support survival and growth of preantral (secondary) follicles from rhesus monkeys to the small antral stage. In this study, a greater percentage of secondary follicles from ovaries of prepubertal and young adult monkeys survived during the 5-week culture period than those from older adult animals. Our previous study demonstrated that follicles isolated during the follicular phase of the menstrual cycle had a higher survival rate than those from the luteal phase, but this may relate to differences in follicle size at onset of culture \( (Xu \ et \ al. \ 2009a) \). The mechanisms resulting in lower survival rate of follicles from older adult monkeys remain unclear, but it is not due to differences in follicle sizes at collection. Unknown factors responsible for the development of preantral follicles may protect them from programmed cell death \( (Orisaka \ et \ al. \ 2006) \). Preantral follicles from older animals may lack these apoptosis inhibiting factors and may not survive in culture. Alternatively, local factors inducing follicle atresia during early follicular development \( (Thompson \ et \ al. \ 2004) \) may increase during aging and result in decreased survival rate of preantral follicles from older animals. This feature of ovarian aging may be relevant to the clinical observation that the oocyte quality and fecundity, either spontaneous or assisted, decline by 30 years of age in premenopausal women \( (Gougeon \ 2005) \).

FSH is a critical hormone for survival of macaque secondary follicles \( (125–225 \mu m \) in diameter) in the alginate-based 3D culture system, regardless of animal age. Recently, \( Kreeger \ et \ al. \ (2005) \) examined the importance of FSH for survival, growth, and cell activity in murine secondary follicles cultured in a similar system. Notably, survival of multilayered follicles \( (150–180 \mu m) \) was markedly improved by exposure to FSH, in a dose-dependent manner, whereas survival of secondary follicles \( (100–130 \mu m) \) was not altered by FSH exposure. The authors distinguished these groups of secondary follicles as FSH dependent versus FSH responsive, as the smaller follicles still exhibited dose-dependent increases in growth, lactate production, and steroid secretion. Thus, the larger macaque secondary follicles appear FSH dependent comparable to those of the mouse, or perhaps even more so since some murine follicles survived in the absence of FSH. Likewise, FSH acts as a survival factor for human preantral follicles during ovarian tissue culture \( (Wright \ et \ al. \ 1999) \). These data are consistent with the evidence that FSH receptors are expressed in preantral follicles of various species, including primates \( (Gougeon \ 1996, \ Findlay \ & \ Drummond \ 1999) \). However, an essential role for FSH in survival, as well as in growth, of preantral follicles in vitro is counter to evidence in vivo that human follicles survive and some grow to the small antral stage in conditions of low-to-nondetectable FSH levels, e.g. in infancy \( (Peters \ 1979) \) and in women during pregnancy \( (Khattab \ & \ Jequier \ 1979) \), in disorders of hypogonadotrophic hypogonadism \( (Goldenberg \ et \ al. \ 1976) \), and selective FSH \( (Rabin \ et \ al. \ 1972) \) or FSH receptor \( (Aittomäki \ et \ al. \ 1996) \) defects. Perhaps, the ovarian milieu/tissue in vivo, in the absence or presence of gonadotropins, can promote survival of small growing follicles.
The surviving macaque secondary follicles could be categorized into three groups (NG, SG, and FG) according to their growth rate, i.e. diameters achieved by culture day 40. This phenomenon was not documented in previous studies on in vitro follicle growth. During similar 3D culture, mouse secondary follicles grew from 200 to 400 μm diameter, reached the antral stage within 8 days, and provided mature oocytes for IVF (Spears et al. 1998, Xu et al. 2006). Any diversity in growth rate among murine follicles may be less evident due to the shorter culture period and smaller increase (doubling) in follicle diameter. In initial studies on human secondary follicles (Xu et al. 2009b), the differences in follicle growth rate may not have been obvious due to small sample size. However, it was observed that 75% of the surviving follicles developed visible antrum, while others remained at multilayered stage through 30 days of culture. The current data suggest that the population of small secondary follicles in the primate ovary at the early follicular phase of the cycle is heterogeneous in their capacity to grow in an FSH-replete milieu. Their growth rate may depend upon their ability to recognize or respond to FSH (Kreeger et al. 2005) or other hormones (e.g. insulin, see below), or to synthesize and respond to other local factors that modulate follicular growth. For example, the heterogeneity of the small preantral follicle pool is supported by the proportion of size-matched preantral follicles in the marmoset ovary that immunolocalized AMH (Thomas et al. 2007).

In addition, the proportion of follicles in the three groups and their growth rates differed as a function of animal age. The NG follicles were observed and exhibited minimal growth in all the three age groups. However, the SG follicles from young adult animals grew faster and reached a larger size than those from prepubertal monkeys. Moreover, the FG follicles were only observed in young adult group. Also during culture with FSH, follicles from young adult animals yielded more healthy GV-intact oocytes with larger diameters than those from prepubertals. The latter observation is consistent with the report that FSH treatment of postmenarcheal rhesus monkeys yielded more mature oocytes with higher development potential during IVF than treatment of juveniles (Yang et al. 2009). In different species, including primates, FSH receptor expression increases during early folliculogenesis, and the receptors remain on granulosa cells of healthy follicles until they become atretic or luteinize (Findlay & Drummond 1999). Again, it is possible that compared with follicles from young adults, the FSH receptor expression and signaling pathway were not fully developed in follicles from prepubertals. The latter observation is consistent with the report that FSH treatment of postmenarcheal rhesus monkeys yielded more mature oocytes with higher development potential during IVF than treatment of juveniles (Yang et al. 2009). In different species, including primates, FSH receptor expression increases during early folliculogenesis, and the receptors remain on granulosa cells of healthy follicles until they become atretic or luteinize (Findlay & Drummond 1999). Again, it is possible that compared with follicles from young adults, the FSH receptor expression and signaling pathway were not fully developed in follicles from prepubertals. The latter observation is consistent with the report that FSH treatment of postmenarcheal rhesus monkeys yielded more mature oocytes with higher development potential during IVF than treatment of juveniles (Yang et al. 2009). In different species, including primates, FSH receptor expression increases during early folliculogenesis, and the receptors remain on granulosa cells of healthy follicles until they become atretic or luteinize (Findlay & Drummond 1999). Again, it is possible that compared with follicles from young adults, the FSH receptor expression and signaling pathway were not fully developed in follicles from prepubertals. The latter observation is consistent with the report that FSH treatment of postmenarcheal rhesus monkeys yielded more mature oocytes with higher development potential during IVF than treatment of juveniles (Yang et al. 2009).
with the 2-cell, 2-gonadotropin model wherein LH receptor signaling promotes A4 production by theca cells, which allows steroidogenic maturation of the follicles by providing A4 as substrate for E2 production in the granulosa cells (McNatty et al. 1980). Elevated P4 levels in the culture media may represent substrate for A4 and E2 production or be a sign of luteinization in follicles with degenerate oocytes (Fauser & Van Heusden 1997). In contrast, an LH effect on steroidogenesis was not observed in the FG follicles. This may be due to the high steroid production prior to the LH addition, which prevented further stimulation. In our previous study, continuous LH exposure during follicle culture promoted E2, but not A4 and P4, production (Xu et al. 2009a). This may result from the desensitizing of LH receptors by continuous LH exposure or by not distinguishing SG from FG follicles. Our current data also indicate that LH had no effect on AMH and VEGF production during follicle culture. Since AMH levels declined to baseline at antrum formation, this transient follicular activity preceded LH exposure at day 30 of culture. In contrast, VEGF production, at least by granulosa cells, appears to reach the capacity in the large preovulatory follicle and is regulated by surge levels of gonadotropin (Ravindranath et al. 1992, Christenson & Stouffer 1997). This study suggests that low nonsurge levels of LH do not promote VEGF production by small antral follicles.

Insulin may regulate various intracellular processes in the follicle such as amino acid transport, lipid metabolism, gene transcription, and protein synthesis (Louhio et al. 2000). Insulin receptors are expressed in mammalian ovaries; its mRNA and protein are located in theca, granulosa, and stromal cells of antral follicles in women (el-Roeiy et al. 1993, Samoto et al. 1993). Insulin receptor gene expression was also detected in in vitro matured oocytes from small antral follicles in monkeys (Zheng et al. 2007). In this study, it was observed that insulin had no effect on follicle survival, but at the higher dose, it promoted further growth of the FG follicles from young adult animals after 3 weeks of culture. This was the time when the follicles reached ≥400 μm in diameter, and the antral cavity started to form. Also, more healthy GV-intact oocytes were retrieved and their sizes tended to be larger in the follicles cultured with high-dose insulin. Our results support previous reports that insulin had a positive effect on ovarian folliculogenesis and oocyte maturation, including human follicular growth in vitro (Wright et al. 1999). However, an in vitro study on murine cumulus–oocyte complexes (COC) indicated that insulin can have profound detrimental effects on oocyte developmental competence (Eppig et al. 1998). Insulin or FSH may also modulate local insulin-like growth factor (IGF) activity, which was suggested to improve the viability of cultured follicles in primates (Louhio et al. 2000). Further studies on the effects of insulin and IGFs, both of which are present at appreciable levels in follicular fluid of primate antral follicles (Brogan et al. 2010), on follicle growth and oocyte quality are warranted in primates.

Follicle growth rate correlated with steroid hormone production in vitro. Unlike NG follicles, the SG and FG follicles produced appreciable levels of A4, E2, and P4 within 5 weeks of culture in the encapsulated 3D system. Media steroid levels first increased when the antrum appeared as similarly reported in mouse follicles in culture (West-Farrell et al. 2009). Since A4 production is generally limited to the theca layer (Tamura et al. 1992), this encapsulated 3D system may support theca cell development or function in macaque, as well as in mouse (Xu et al. 2006), follicles. Interestingly, this steroidogenic process was observed in follicles cultured with high-dose, but not low-dose, insulin. In mammals, insulin has been shown to promote theca and granulosa cell steroidogenesis (Barbieri et al. 1984, Erickson et al. 1990, Langhout et al. 1991). Thus, insulin/IGFs may be an important regulator of primate theca–granulosa differentiation and steroidogenesis in vitro.
Despite exposure to the same dose of insulin, cultured follicles from prepubertal and older adult animals displayed lower ovarian steroid levels in the media. This may be due to lower FSH sensitivity of the follicles as described above (Davoren & Hsueh 1984). Follicle growth may be delayed, and follicle function may be impaired in prepubertal and older adults if insulin-enhanced FSH-stimulated steroidogenesis is lacking.

This study provided the first quantitative assessment of AMH production by isolated primate preantral follicles. AMH production by early preantral follicles correlated positively with growth rate, i.e. the FG follicles produced greater levels than the SG follicles, with lesser production by the NG follicles. AMH production displayed a transient increase at culture week 2 and decreased by antrum formation. Previous studies localized AMH mRNA or protein to granulosa cells of preantral and small antral follicles, which diminished in the subsequent stages of follicle development; expression was also absent in atretic follicles and the corpus luteum (Durlinger et al. 2002, Weenen et al. 2004, Thomas et al. 2007). Thus, early AMH production may be a potential marker for predicting further development of cohorts of preantral follicles with different growth rates during culture, prior to any differences in follicle diameter and steroidogenic capacity. Efforts continue attempting to elucidate the role of AMH on follicular function (Visser et al. 2007, La Marca et al. 2009). The addition of AMH into cultures of human or rat ovarian cortical strips, improved the recruitment, survival, and growth of primordial follicles (McGee et al. 2001, Schmidt et al. 2005) suggesting a stimulatory role of AMH on very early follicular development. However, a direct role for AMH on primary or secondary follicle function has yet to be demonstrated. AMH may not only be a marker, but also an important local factor regulating follicle growth in encapsulated 3D culture system.

This study also demonstrated, for the first time, that small antral follicles in primates produce VEGF. Media VEGF levels increased at culture week 3 during antrum formation, correlated with growth rate, and were greatest in the FG follicles. The pattern is consistent with previous in vivo studies that VEGF mRNA (Ravindranath et al. 1992) and protein (Yamamoto et al. 1997) were expressed in the theca cells of antral follicles and granulosa cells nearest the oocyte in the preovulatory follicle of primates, but not in the granulosa cells of primordial and preantral follicles. VEGF likely plays an angiogenic role during antrum development when the theca layer acquires a vascular sheath to provide an increased supply of gonadotropins, growth factors, oxygen, and steroid precursors to the growing follicle (Stouffer et al. 2001). Increased VEGF production by the SG and FG follicles in encapsulated 3D culture may indicate achievement of a size and maturation state in the follicle, at the antral stage, that requires vascularization to achieve further development in vivo with additional substrates and release of hormones. Besides an angiogenic action, VEGF may also be a cytoprotective factor in the extravascular granulosa cell compartment. Co-expression of VEGF and its receptor reportedly protects bovine granulosa cells from apoptotic cell death and follicle atresia (Greenaway et al. 2004). VEGF may also promote nuclear and cytoplasmic maturation of bovine oocytes in vitro (Luo et al. 2002). Given that increasing follicle diameter correlated positively with increased VEGF production, VEGF may play a role during follicle development and/or be a marker of follicle quality in vitro.

In addition to appreciable number of degenerate oocytes, the healthy oocytes within the SG or FG follicles after 40 days of culture did not respond to rhCG treatment with reinitiation of meiotic maturation and remained at the GV-intact stage. In order to acquire meiotic competence, the oocytes need to grow and reach a stage at which they are able to activate cyclin-dependent protein kinase (CDC2) and MAP kinases (Miyano & Manabe 2007). Chromatin configurations and meiotic competence of oocytes are related to follicular diameter as reported in rodents (Erickson & Sorensen 1974, Iwamatsu & Yanagimachi 1975) and nonstimulated rhesus monkeys. Limited data from rhesus monkeys suggest that 56% of oocytes from large antral follicles (>100 µm in diameter) completed maturation, whereas few (9%) oocytes from small antral follicles (200–450 µm in diameter) were competent to mature in vitro (Schramm et al. 1993). The current culture systems need to be improved to promote further follicle growth, as well as the health and further growth of the oocytes to reach the size of those that mature in vivo, which is over 100 µm in diameter in macaque (Buse et al. 2008). Gap junctional communication within the COC matrix is likely critical for oocyte metabolism, maturation, and meiotic progression in primates, as in other species (Kimura et al. 2007). However, the cumulus–oocyte communication could be compromised by the current milieu and extended interval of encapsulated 3D culture, such that oocytes cannot achieve the competence to undergo meiotic or cytoplasmic maturation. Future studies on the characteristics of COC of antral follicles derived from secondary follicles during encapsulated 3D culture are warranted.

Thus, advances in an encapsulated 3D culture of nonhuman primate secondary follicles to the small antral stage provide a model to study indices of follicle development with individual follicles. This model offers an opportunity to characterize the endocrine and paracrine function of primate follicles that influence follicle growth and oocyte maturation, with relevance to translational efforts to grow human follicles in vitro. By achieving the goal of producing competent oocytes, in vitro follicle maturation may offer a means to enhance fertility preservation options in women.
Materials and Methods

Animals and ovary collection

The general care and housing of rhesus macaques were provided by the Division of Animal Resources at the Oregon National Primate Research Center (ONPRC). Animals, different from those used in our previous study (Xu et al. 2009a), were pair caged in a temperature-controlled (22 °C) light-regulated 12 h light:12 h darkness room. Diet consisted of Purina monkey chow (Ralston-Purina, Richmond, IN, USA) provided twice a day, supplemented with fresh fruit or vegetables once a day and water made available ad libitum. Animals were treated according to the National Institutes of Health Guide for the Care and Use of Laboratory Animals and protocols approved by the ONPRC Institutional Animal Care and Use Committee.

Young adult female rhesus macaques (n = 3; 4–11 years of age) exhibiting regular menstrual cycles of ~28 days were evaluated daily for menstruation with the first day of menses being termed day 1 of the cycle. Ovariectomies were conducted on anesthetized monkeys by laparoscopy at early pubertal (Duffy & Stouffer 2002). Ovaries were also obtained from follicular phase, days 1–3 of the cycle, as described previously (Xu et al. 2009a). Ovaries were immediately transferred into HEPES-buffered holding media (as described above) using 31-gauge needles, and the secondary follicles with diameters of 125–225 μm were transferred into 5 ml holding media (as described above) using 31-gauge needles, and the droplets were crosslinked in 50 mM CaCl₂. Cortex was cut into 1 mm cortical strips and incubated and the secondary follicles with diameters of 125–225 μm were transferred into 5 ml holding media (as described above) containing sterile sodium alginate (FMC BioPolymers, Philadelphia, PA, USA) in PBS (137 mM NaCl, 10 mM phosphate, 2.7 mM KCl, 1.8 mM CaCl₂, and 0.25% (w/v) human serum protein supplement (SPS; CooperSurgical, Inc.) and 10 μg/ml gentamicin (Sigma–Aldrich).

Follicle isolation, encapsulation, and culture

Follicle isolation and encapsulation were carried out as described previously (Xu et al. 2009a). Briefly, the ovarian cortex was cut into 2 × 2 × 1 mm cortical strips and incubated in 6 ml holding media (as described above) containing 275 U/ml collagenase type 1 and 585 U/ml DNase I (Worthington Biochemical Corp., Lakewood, NJ, USA) at 37 °C for 30 min. Follicles were mechanically isolated in the holding media (as described above) using 31-gauge needles, and the secondary follicles with diameters of 125–225 μm that displayed the following characteristics were selected for encapsulation: i) no clear antral cavity, ii) an intact basement membrane with attached stroma, and iii) a visible oocyte that was round and centrally located within the follicle.

Follicles were transferred individually into 5 μl 0.25% (w/v) sterile sodium alginate (FMC BioPolymers, Philadelphia, PA, USA) in PBS (137 mM NaCl, 10 mM phosphate, 2.7 mM KCl, Invitrogen), and the droplets were crosslinked in 50 mM CaCl₂ and 140 mM NaCl solution for 2 min. Each alginate-encapsulated follicle was then transferred into individual wells of 48-well plates containing 300 μl of minimum essential medium (Invitrogen) supplemented with 0.3% SPS, 1 mg/ml bovine fetuin, 5 μg/ml transferrin, and 5 ng/ml sodium selenite (Sigma–Aldrich).

Encapsulated follicles were cultured at 37 °C in 5% (v/v) CO₂ in an atmosphere in the absence or presence of 500 μl/ml rhFSH (NV Organon, Oss, The Netherlands) and 0.05 or 5 μg/ml insulin (low or high dose, Sigma–Aldrich) for 30 days. Thereafter, half of the follicles in each culture condition received 10 mIU/ml rhLH (Ares-Serono, Inc., Randolph, MA, USA) in the media from days 30–40. Follicles that reached the antral stage were treated with 100 ng/ml rhCG (Merck Serono) for 34 h. Oocytes were then retrieved to determine competence for meiotic maturation. Half of the culture media (150 μl) was collected and replaced every other day and stored at −20 °C. The three media samples from each culture week were assigned to ovarian steroids, AMH, and VEGF assays.

Follicle survival, growth, and oocyte maturation

Follicle survival, diameter, and antrum formation were assessed weekly using an Olympus CK40 inverted microscope and an Olympus DP11 digital camera (Olympus Imaging America, Inc., Center Valley, PA, USA) as described previously (Xu et al. 2009a). Follicles were measured from the outer layer of cells, which included a measurement at the widest diameter of the follicle and a second measurement perpendicular to the first. The mean of these values was then calculated and reported as the follicle diameter. Any clumps of presumptive discontinuous stromal cells (Fig. 1B–D) were not included. Follicles were considered undergoing atresia if the oocyte was dark or not surrounded by a layer of granulosa cells, the granulosa cells became dark and fragmented, or the diameter of the follicle decreased. Follicle photographs were imported into ImageJ 1.42 software (National Institutes of Health, Bethesda, MD, USA), and the diameter of each follicle was measured. Retrieved oocytes were also photographed, and oocyte diameters were measured using the same camera and software as described above.

Ovarian steroids, AMH, and VEGF assays

Media collected during culture weeks 1–5 were analyzed for E₂ and P₄ concentrations by the Endocrine Technology Support Core at the ONPRC using an Immulite 2000, a chemiluminescence-based automatic clinical platform (Siemens Healthcare Diagnostics, Deerfield, IL, USA), according to the manufacturers’ protocol as reported previously (Xu et al. 2009a). Media A4 levels were measured by RIA using a DSL-3800 kit (Diagnostic Systems Laboratories, Inc., Webster, TX, USA). The assay was validated for parallelism with blank culture media containing a known concentration of A4, and with media used in culture with monkey follicles. The sensitivity of the assay was 0.1 ng/ml for 50 μl sample. The standard curve of the assay ranged 0.1–10 ng/ml. The coefficient of variation (CV) values for standard points (in duplicate) at 0, 0.1, 0.3, 1.0, 3.0, and 10.0 ng/ml in the assays were 0.7, 0.5, 1.4, 1.8, 0.1, and 1.5% respectively.

Media collected during culture weeks 1–5 were analyzed for AMH concentrations by ELISA using a DSL-10-14400 kit (Diagnostic Systems Laboratories, Inc.) based on the manufacturers’ instructions (Fréour et al. 2007). The assay was validated for parallelism with blank culture media containing a known concentration of AMH, and with media used in culture with monkey follicles. The sensitivity of the assay was 0.05 ng/ml for 20 μl sample. The standard curve of the assay ranged 0.05–14 ng/ml. The CV values for standard points (in duplicate) at 0, 0.05, 0.1, 0.26, 2.0, 7.8, and 14.0 ng/ml in the assays were...
Statistical analysis

Statistical significance was analyzed by SigmaPlot 11 software (SPSS, Inc., Chicago, IL, USA) using a two-way ANOVA with repeated measures or one-way ANOVA followed by the Student Newman–Keuls post hoc test for single time points. Differences were considered significant at \( P<0.05 \), and values are presented as mean \( \pm \) S.E.M.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding

This work was supported by the National Institutes of Health (grant number NIH UL1 RR024926 (R01-HD058294, PL1-EB008542), U54-HD18185, NCRR-RR00163, HD07133, D43TW000668-Fogarty).

Acknowledgements

We are grateful to the members of the Division of Animal Resources, the Endocrine Technology Support Core, the Assisted Reproductive Technology/Embryonic Stem Cell Support Core, and the Colony Demographics and Informatics Unit at the ONPRC, as well as to Drs Min Xu, Lonnie Shea, and Teresa Woodruff at the Northwestern University, for their valuable expertise and technical assistance. We thank N V Organon for providing the rhFSH.

References


Christensen LK & Stouffer RL 1997 Follicle-stimulating hormone and luteinizing hormone/chorionic gonadotropin stimulation of vascular endothelial growth factor production by macaque granulosa cells from pre- and periovulatory follicles. Journal of Clinical Endocrinology and Metabolism 82 2135–2142. (doi:10.1210/jc.82.7.2135)


Iwamatsu T & Yanagimachi R 1993 Follicle stimulating hormone levels in serum of pregnant female chimpanzees during the early and late pregnancy periods. *Journal of Reproduction and Fertility* 100 293–298. (doi:10.1530/jr.0.1350589)


Schramm RD, Tennier MT, Boatman EE & Bavister BD 1993 Chromatin configurations and meiotic competence of oocytes are related to follicular diameter in nonstimulated rhesus monkeys. *Endocrinology* 129 349–356. (doi:10.1210/endo.48.2.349)


Received 22 June 2010
First decision 19 August 2010
Accepted 19 August 2010